

## Fixation Requirements for Surgical Specimen Submission v2.1

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### 148355.278 Fixation Requirements for Surgical Specimen Submission

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#### Comments for version 2.1

Initial version

#### Approval and Periodic Review Signatures

Type	Description	Date	Version	Performed By	Notes
Approval	Lab Director	5/16/2022	2.1	John Fisk MD Clinical Laboratory Director (M08480)	
Approval	Lab Director	5/12/2022	2.1	Simha Sastry MD Clinical Laboratory Director (M06625)	
Approval	Lab Director	5/11/2022	2.1	Valerie Bush PhD Clinical Laboratory Director (M05512)	
Approval	Lab Director	5/10/2022	2.1	Samantha Davenport MD Service Line Chief (M03764)	
Approval	Lab Director	5/8/2022	2.1	Ghazala Nathu MD Clinical Laboratory Director (S00134)	
Approval	Lab Director	5/2/2022	2.1	Timothy Chapman MD Clinical Laboratory Director (M11669)	

#### Version History

Version	Status	Type	Date Added	Date Effective	Date Retired
2.1	Approved and Current	Initial version	4/22/2022	5/16/2022	Indefinite

## Fixation Requirements for Surgical Specimen Submission

- I. During regular working days from 0700-1630 hours:
  - A. The following specimens should be delivered fresh to Surgical Pathology. Small specimens should be placed in saline moistened gauze. Brain biopsies should be placed in a miniscule amount of saline in a sterile container.
    1. All large specimens for gross examination or photography.  
(E.g. stomach, larynx, placenta, colon, limbs, pancreas)
    2. Tissues for culture as well as histology (to be routed to Microbiology first)
    3. Any tissue for frozen section
    4. Any tissue for operating room consultation or open and return.
    5. Lymph node (may also be placed in saline moistened gauze).
  - B. Handling instructions for the following specimens, may be obtained from the gross room staff prior to removal.
    1. Muscle biopsy (1.5 cm x 1.5 cm piece submitted fresh within 15 minutes.)
    2. Skin biopsy for immunofluorescence (Michel's transport media).
    3. Renal biopsy (divided in three portions and submitted respectively in Glutaraldehyde, Zeus, and Formalin.)
    4. Any biopsy for special immunohistochemistry.
    5. Genetic testing (three, 2.0cm sterile tissue samples in RPMI)
    6. Small Fiber Neuropathy (two, 3 mm samples in Zamponi's fixative)
  - C. All other tissue should be immediately placed in 10% neutral buffer formalin and delivered to the Laboratory.

II. During non-operating hours:

A. Tissue usually sent fresh should be treated in the following manner:

1. All large specimens should be placed in the Surgical Pathology refrigerator.
2. All tissue for culture as well as histology should be sent to the Microbiology Laboratory. The Microbiology Technologist will process the specimen and refrigerate the remaining tissue for histology.
3. All other fresh specimens will require consultation by the On-call-Pathologist.

B. Handling instructions for the following specimens, MUST be obtained from the on-call-Pathologist prior to removal.

1. Muscle biopsy (two, 1.5cm long pieces submitted fresh within 15 minutes.)
2. Skin biopsy for immunofluorescence (Michel's transport media.)
3. Renal biopsy (divided in three portions and submitted respectively in Glutaraldehyde, Zeus, and Formalin.)
4. Any biopsy for special immunohistochemistry.
5. Genetic testing (three, 2.0cm sterile tissue samples in RPMI)
6. Small Fiber Neuropathy (two, 3 mm samples in Zamponi's fixative)

C. All other specimens should be placed in 10% buffered formalin and delivered to the laboratory.